

- Rigopoulos, N., Thomou, E., Kouloumpis, A., Lamprou, E.R., Petropoulea, V., Gournis, D., Poulis, E., Karantonis, H.C. & Giaouris, E. 2018. Optimization of Silver Nanoparticle Synthesis by Banana Peel Extract Using Statistical Experimental Design, and Testing of their Antibacterial and Antioxidant Properties. *Current Pharmaceutical Biotechnology*. 20(10):858–873.
- Rodrigues, M.E., Henriques, M. & Silva, S. 2016. Disinfectants to fight oral Candida biofilms. *Advances in Experimental Medicine and Biology* Vol. 931. Springer New York: 83–93.
- Rodrigues, C.F., Rodrigues, M.E. & Henriques, M.C.R. 2019. Promising Alternative Therapeutics for Oral Candidiasis. *Current Medicinal Chemistry*. 26(14):2515–2528.
- Rosa-garcía, S.C. De, Martínez-torres, P., Gómez-cornelio, S., Corral-aguado, M.A., Quintana, P. & Gómez-ortíz, N.M. 2018. Antifungal Activity of ZnO and MgO Nanomaterials and Their Mixtures against *Colletotrichum gloeosporioides* Strains from Tropical Fruit. *Journal of Nanomaterials*. 2018(3):1–9.
- Rozman, N.A.S., Yenn, T.W., Ring, L.C., Nee, T.W., Hasanolbasori, M.A. & Abdullah, S.Z. 2019. Potential antimicrobial applications of chitosan nanoparticles (ChNPS). *Journal of Microbiology and Biotechnology*. 29(7):1009–1013.
- Ruddaraju, L.K., Pammi, S.V.N., Guntuku, G. Sankar, Padavala, V.S. & Kolapalli, V.R.M. 2020. A review on anti-bacterials to combat resistance: From ancient era of plants and metals to present and future perspectives of green nano technological combinations. *Asian Journal of Pharmaceutical Sciences*. 15(1):42–59.
- Sacarlal, J. & Denning, D.W. 2018. Estimated burden of serious fungal infections in Mozambique. *Journal of Fungi*. 4(3):75.
- Saffarpour, M., Rahmani, M., Tahriri, M. & Peymani, A. 2016. Antimicrobial and bond strength properties of a dental adhesive containing zinc oxide nanoparticles. *Brazilian Journal of Oral Sciences*. 15(1):66–69.
- Samiei, M., Torab, A., Hosseini, O., Abbasi, T., Abdollahi, A.A. & Divband, B. 2018. Antibacterial effect of two nano zinc oxide gel preparations compared to calcium hydroxide and chlorhexidine mixture. *Iranian Endodontic Journal*. 13(3):305–311.
- Sánchez-López, E., Gomes, D., Esteruelas, G., Bonilla, L., Laura Lopez-Machado, A., Galindo, R., Cano, A., Espina, M., Ettcheto, M., Camins, A., Silva, A.M., Durazzo, A., Santini, A., Garcia, M.L. & Souto, E.B. 2020. Metal-Based Nanoparticles as Antimicrobial Agents: An Overview. *Nanomaterials*. 10:292.
- Sanders, W.C. 2018. *Basic principles of nanotechnology*. CRC Press: 1–78.
- Sangeetha, G., Rajeshwari, S. & Venckatesh, R. 2011. Green synthesis of zinc oxide nanoparticles by aloe barbadensis miller leaf extract: Structure and optical properties. *Materials Research Bulletin*. 46(12):2560–2566.
- Santhoshkumar, J., Kumar, S.V. & Rajeshkumar, S. 2017. Synthesis of zinc oxide nanoparticles using plant leaf extract against urinary tract infection pathogen. *Resource-Efficient Technologies*. 3(4):459–465.
- Santos, J.S., Deolindo, C.T.P., Esmerino, L.A., Genovese, M.I., Fujita, A., Marques, M.B., Rosso, N.D., Dagher, H., Valse, A. C. and Granato, D. 2016. Effects of time and extraction temperature on phenolic composition and functional properties of red rooibos (*Aspalathus linearis*). *Food Research International*. 89:476–487.

- Sasaki, M., Nishida, N. & Shimada, M. 2018. A Beneficial Role of Rooibos in Diabetes Mellitus: A Systematic Review and Meta-Analysis. *Molecules*. 23(4):839.
- Saubolle, M.A. & Hoepflich, P.D. 1978. Disk agar diffusion susceptibility testing of yeasts. *Antimicrobial Agents and Chemotherapy*. 14(4):517–530.
- Scheibler, E., da Silva, R.M., Leite, C.E., Campos, M.M., Figueiredo, M.A., Salum, F.G. & Cherubini, K. 2018. Stability and efficacy of combined nystatin and chlorhexidine against suspensions and biofilms of *Candida albicans*. *Archives of Oral Biology*. 89:70–76.
- Schiefersteiner, M., Bichsel, D., Rücker, M. & Valdec, S. 2019. Antimycotics in dental routine – an update. *Swiss dental journal*. 129(5):403–405.
- Schwartz, I.S., Boyles, T.H., Kenyon, C.R., Hoving, J.C., Brown, G.D. & Denning, D.W. 2019. The estimated burden of fungal disease in South Africa. *South African Medical Journal*. 109(11):885.
- Selim, Y.A., Azb, M.A., Ragab, I. & H. M. Abd El-Azim, M. 2020. Green Synthesis of Zinc Oxide Nanoparticles Using Aqueous Extract of *Deverra tortuosa* and their Cytotoxic Activities. *Scientific Reports*. 10(1):1–9.
- Seneviratne, C.J., Jin, L. & Samaranayake, L.P. 2008. Biofilm lifestyle of *Candida*: A mini review. *Oral Diseases* 14 (7) p.582–590.
- Seong, M. & Lee, D.G. 2018. Reactive oxygen species-independent apoptotic pathway by gold nanoparticles in *Candida albicans*. *Microbiological Research*. 207:33–40.
- Sevinç, B.A. & Hanley, L. 2010. Antibacterial activity of dental composites containing zinc oxide nanoparticles. *Journal of Biomedical Materials Research - Part B Applied Biomaterials*. 94(1):22–31.
- Shaikh, S., Nazam, N., Rizvi, S.M.D., Ahmad, K., Baig, M.H., Lee, E.J. & Choi, I. 2019. Mechanistic Insights into the Antimicrobial Actions of Metallic Nanoparticles and Their Implications for Multidrug Resistance. *International journal of molecular sciences*. 20(10):2468.
- Sharma, D., Sabela, M.I., Kanchi, S., Mdluli, P.S., Singh, G., Stenström, T.A. & Bisetty, K. 2016. Biosynthesis of ZnO nanoparticles using *Jacaranda mimosifolia* flowers extract: Synergistic antibacterial activity and molecular simulated facet specific adsorption studies. *Journal of Photochemistry and Photobiology B: Biology*. 162:199–207.
- Sharma, A. 2018. Oral candidiasis: An opportunistic infection: A review Amrit Sharma. [Online], Available: www.oraljournal.com [Accessed: 2020, May 20].
- Sharma, A. 2019. Oral Candidiasis: An Opportunistic infection-A Review. *International Journal of Applied Dental Sciences*. 5(1):23–27.
- Sharmila, G., Thirumarimurugan, M. & Muthukumaran, C. 2019. Green synthesis of ZnO nanoparticles using *Tecoma castanifolia* leaf extract: Characterization and evaluation of its antioxidant, bactericidal and anticancer activities. *Microchemical Journal*. 145:578–587.
- Shashirekha, G., Jena, A. & Mohapatra, S. 2017. Nanotechnology in Dentistry: Clinical Applications, Benefits, and Hazards. *Compendium of continuing education in dentistry*. 38(5):e1–e4.
- Shayegan Mehr, E., Sorbiun, M., Ramazani, A. & Taghavi Fardood, S. 2018. Plant-mediated synthesis of zinc oxide and copper oxide nanoparticles by using *Ferulago angulata*

- (schlecht) boiss extract and comparison of their photocatalytic degradation of Rhodamine B (RhB) under visible light irradiation. *Journal of Materials Science: Materials in Electronics*. 29(2):1333–1340.
- Shekhawat, M.S. & Manokari, M. 2014. Biogenesis of Zinc oxide Nanoparticles using *Morinda pubescens*. *International Journal of BioEngineering and Technology*.5(1):1–6.
- Sheppard, D.C. & Howell, P.L. 2016. Biofilm exopolysaccharides of pathogenic fungi: Lessons from bacteria. *Journal of Biological Chemistry*. 291(24):12529–12537.
- Shin, J., Prabhakaran, V.S. & Kim, K.-S. 2018. The multi-faceted potential of plant-derived metabolites as antimicrobial agents against multidrug-resistant pathogens. *Microbial Pathogenesis*. 116:209–214.
- Shoeb, M., Singh, B.R., Khan, J.A., Khan, W., Singh, B.N., Singh, H.B. & Naqvi, A.H. 2013. ROS-dependent anticandidal activity of zinc oxide nanoparticles synthesized by using egg albumen as a biotemplate. *Advances in Natural Sciences: Nanoscience and Nanotechnology*. 4(3):035015.
- Shukla, S.K. & Rao, T.S. 2017. An Improved Crystal Violet Assay for Biofilm Quantification in 96-Well Microtitre Plate. bioRxiv 100214
- da Silva, B.L., Abuçafy, M.P., Manaia, E.B., Junior, J.A.O., Chiari-Andréo, B.G., Pietro, R.C.L.R. & Chiavacci, L.A. 2019a. Relationship between structure and antimicrobial activity of zinc oxide nanoparticles: An overview. *International Journal of Nanomedicine*. 14:9395–9410.
- da Silva, B., Caetano, B.L., Chiari-Andréo, B.G., Pietro, R.C.L.R. & Chiavacci, L.A. 2019b. Increased antibacterial activity of ZnO nanoparticles: Influence of size and surface modification. *Colloids and Surfaces B: Biointerfaces*. 177:440–447.
- Siddiqi, K.S. & Husen, A. 2016. Fabrication of Metal Nanoparticles from Fungi and Metal Salts: Scope and Application. *Nanoscale Research Letters*. 11(1):1–15.
- Siddiqi, K.S., ur Rahman, A., Tajuddin & Husen, A. 2018. Properties of Zinc Oxide Nanoparticles and Their Activity Against Microbes. *Nanoscale Research Letters*. 13:141.
- Simpson, D. 1998. Buchu - South Africa's amazing herbal remedy. *Scottish Medical Journal*. 43(6):189–191.
- Singh, A., Verma, R., Murari, A. & Agrawal, A. 2014. Oral candidiasis: An overview. *Journal of Oral and Maxillofacial Pathology*. 18(5):81–85.
- Singh, B., Singh, J.P., Kaur, A. & Singh, N. 2016. Bioactive compounds in banana and their associated health benefits - A review. *Food Chemistry*. 206:1–11.
- Singh, J., Vishwakarma, K., Ramawat, N., Rai, P., Singh, V.K., Mishra, R.K., Kumar, V., Tripathi, D.K., *et al.* 2019. Nanomaterials and microbes' interactions: a contemporary overview. *3 Biotech*. 9(3):68.
- Sirelkhatim, A., Mahmud, S., Seeni, A., Kaus, N.H.M., Ann, L.C., Bakhori, S.K.M., Hasan, H. & Mohamad, D. 2015. Review on zinc oxide nanoparticles: Antibacterial activity and toxicity mechanism. *Nano-Micro Letters*. 7(3):219–242.
- Siriyong, T., Srimanote, P., Chusri, S., Yingyongnarongkul, B.E., Suaisom, C., Tipmanee, V. & Voravuthikunchai, S.P. 2017. Conessine as a novel inhibitor of multidrug efflux pump systems in *Pseudomonas aeruginosa*. *BMC Complementary and Alternative Medicine*. 17(1):1–7.

- Skupien, J.A., Valentini, F., Boscato, N. & Pereira-Cenci, T. 2013. Prevention and treatment of *Candida* colonization on denture liners: A systematic review. *Journal of Prosthetic Dentistry*. 110(5):356–362.
- Slot, D.E., Berchier, C.E., Addy, M., Van der Velden, U. & Van der Weijden, G.A. 2014. The efficacy of chlorhexidine dentifrice or gel on plaque, clinical parameters of gingival inflammation and tooth discoloration: A systematic review. *International Journal of Dental Hygiene*. 12(1):25–35.
- Smith, C. & Swart, A. 2018. *Aspalathus linearis* (Rooibos) – a functional food targeting cardiovascular disease. *Food & Function*. 9(10):5041–5058.
- Somu, P. & Paul, S. 2019. A biomolecule-assisted one-pot synthesis of zinc oxide nanoparticles and its bioconjugate with curcumin for potential multifaceted therapeutic applications. *New Journal of Chemistry*. 43(30):11934–11948.
- Sorbiun, M., Shayegan Mehr, E., Ramazani, A. & Taghavi Fardood, S. 2018. Green Synthesis of Zinc Oxide and Copper Oxide Nanoparticles Using Aqueous Extract of Oak Fruit Hull (Jaft) and Comparing Their Photocatalytic Degradation of Basic Violet 3. *International Journal of Environmental Research*. 12(1):29–37.
- Sportelli, M.C., Longano, D., Bonerba, E., Tantillo, G., Torsi, L., Sabbatini, L., Cioffi, N. & Ditaranto, N. 2020. Electrochemical preparation of synergistic nanoantimicrobials. *Molecules*. 25(1):49.
- Stan, M., Popa, A., Toloman, D., Dehelean, A., Lung, I. & Katona, G. 2015. Enhanced photocatalytic degradation properties of zinc oxide nanoparticles synthesized by using plant extracts. *Materials Science in Semiconductor Processing*. 39:23–29.
- Stan, M., Popa, A., Toloman, D., Silipas, T.D. & Vodnar, D.C. 2016. Antibacterial and antioxidant activities of ZnO nanoparticles synthesized using extracts of *Allium sativum*, *Rosmarinus officinalis* and *Ocimum basilicum*. *Acta Metallurgica Sinica (English Letters)*. 29(3):228–236.
- Stepanović, S., Vuković, D., Dakić, I., Savić, B. & Švabić-Vlahović, M. 2000. A modified microtiter-plate test for quantification of staphylococcal biofilm formation. *Journal of Microbiological Methods*. 40(2):175–179.
- Van Strydonck, D.A.C., Slot, D.E., Van Der Velden, U. & Van Der Weijden, F. 2012. Effect of a chlorhexidine mouthrinse on plaque, gingival inflammation and staining in gingivitis patients: A systematic review. *Journal of Clinical Periodontology*. 39(11):1042–1055.
- Sun, Q., Li, J. & Le, T. 2018. Zinc Oxide Nanoparticle as a Novel Class of Antifungal Agents: Current Advances and Future Perspectives. *Journal of Agricultural and Food Chemistry*. 66(43):11209–11220.
- Sundrarajan, M., Ambika, S. & Bharathi, K. 2015. Plant-extract mediated synthesis of ZnO nanoparticles using *Pongamia pinnata* and their activity against pathogenic bacteria. *Advanced Powder Technology*. 26(5):1294–1299.
- Supranoto, S., Slot, D., Addy, M. & Van der Weijden, G. 2015. The effect of chlorhexidine dentifrice or gel versus chlorhexidine mouthwash on plaque, gingivitis, bleeding and tooth discoloration: a systematic review. *International Journal of Dental Hygiene*. 13(2):83–92.
- Suresh, D., Nethravathi, P.C., Udayabhanu, Rajanaika, H., Nagabhushana, H. & Sharma, S.C. 2015. Green synthesis of multifunctional zinc oxide (ZnO) nanoparticles using *Cassia fistula* plant extract and their photodegradative, antioxidant and antibacterial activities.

- Suresh, J., Pradheesh, G., Alexramani, V., Sundrarajan, M. & Hong, S.I. 2018. Green synthesis and characterization of zinc oxide nanoparticle using insulin plant (*Costus pictus D. Don*) and investigation of its antimicrobial as well as anticancer activities. *Advances in Natural Sciences: Nanoscience and Nanotechnology*. 9(1):015008.
- Susewind, S., Lang, R. & Hahnel, S. 2015. Biofilm formation and *Candida albicans* morphology on the surface of denture base materials. *Mycoses*. 58(12):719–727.
- Sydnés, M. 2014. One-Pot Reactions: A Step Towards Greener Chemistry. *Current Green Chemistry*. 1(3):216–226.
- Taff, H.T., Nett, J.E. & Andes, D.R. 2012. Comparative analysis of *Candida* biofilm quantitation assays. *Medical Mycology*. 50(2):214–218.
- Tariq, S., Wani, S., Rasool, W., Shafi, K., Bhat, M.A., Prabhakar, A., Shalla, A.H. & Rather, M.A. 2019. A comprehensive review of the antibacterial, antifungal and antiviral potential of essential oils and their chemical constituents against drug-resistant microbial pathogens. *Microbial Pathogenesis*. 134:103580.
- Tavassoli Hojati, S., Alaghemand, H., Hamze, F., Ahmadian Babaki, F., Rajab-Nia, R., Rezvani, M.B., Kaviani, M. & Atai, M. 2013. Antibacterial, physical and mechanical properties of flowable resin composites containing zinc oxide nanoparticles. *Dental Materials*. 29(5):495–505.
- Thema, F.T., Manikandan, E., Dhlamini, M.S. & Maaza, M. 2015a. Green synthesis of ZnO nanoparticles via *Agathosma betulina* natural extract. *Materials Letters*. 161:124–127.
- Thema, F.T., Beukes, P., Gurib-Fakim, A. & Maaza, M. 2015b. Green synthesis of Monteponite CdO nanoparticles by *Agathosma betulina* natural extract. *Journal of Alloys and Compounds*. 646:1043–1048.
- Thema, F.T., Manikandan, E., Gurib-Fakim, A. & Maaza, M. 2016. Single phase Bunsenite NiO nanoparticles green synthesis by *Agathosma betulina* natural extract. *Journal of Alloys and Compounds*. 657:655–661.
- Tiew, P.Y., Mac Aogain, M., Ali, N.A.B.M., Thng, K.X., Goh, K., Lau, K.J.X. & Chotirmall, S.H. 2020. The Mycobiome in Health and Disease: Emerging Concepts, Methodologies and Challenges. *Mycopathologia*. 185(2):207–231.
- Tobal, I.E., Roncero, A.M., Moro, R.F., Díez, D. & Marcos, I.S. 2020. Antibacterial natural halimanes: Potential source of novel antibiofilm agents. *Molecules*. 25(7):1707.
- Tournu, H. & Van Dijck, P. 2012. *Candida* biofilms and the host: Models and new concepts for eradication. *International Journal of Microbiology*. 2012.
- Treacy, M.M.J. & Higgins, J.B. 2007. Collection of Simulated XRD Powder Patterns for Zeolites Fifth (5th) Revised Edition. Elsevier:1-387.
- Tsui, C., Kong, E.F. & Jabra-Rizk, M.A. 2016. Pathogenesis of *Candida albicans* biofilm. *Pathogens and disease*. 74(4):1-13.
- Tufa, T.B. & Denning, D.W. 2019. The burden of fungal infections in Ethiopia. *Journal of Fungi*. 5(4):109.
- Turner, M., Jahangiri, L. & Ship, J.A. 2008. Hyposalivation, xerostomia and the complete denture: A systematic review. *The Journal of the American Dental Association*. 139(2):146–150.

- Umar, H., Kavaz, D. & Rizaner, N. 2019. Biosynthesis of zinc oxide nanoparticles using *Albizia lebbbeck* stem bark, and evaluation of its antimicrobial, antioxidant, and cytotoxic activities on human breast cancer cell lines. *International Journal of Nanomedicine*. 14:87–100.
- Uppuluri, P. & Chaffin, W.L. 2007. Defining *Candida albicans* stationary phase by cellular and DNA replication, gene expression and regulation. *Molecular Microbiology*. 64(6):1572–1586.
- Vallet-Regí, M., González, B. & Izquierdo-Barba, I. 2019. Nanomaterials as promising alternative in the infection treatment. *International Journal of Molecular Sciences*. 20(15):3806.
- Vanathi, P., Rajiv, P., Narendhran, S., Rajeshwari, S., Rahman, P.K.S.M. & Venckatesh, R. 2014. Biosynthesis and characterization of Phyto mediated zinc oxide nanoparticles: A green chemistry approach. *Materials Letters*. 134:13–15.
- Vargas-Reus, M.A., Memarzadeh, K., Huang, J., Ren, G.G. & Allaker, R.P. 2012. Antimicrobial activity of nanoparticulate metal oxides against peri-implantitis pathogens. *International Journal of Antimicrobial Agents*. 40(2):135–139.
- Verma, D., Garg, P.K. & Dubey, A.K. 2018. Insights into the human oral microbiome. *Archives of Microbiology*. 200(4):525–540.
- Versiani, M.A., Abi Rached-Junior, F.J., Kishen, A., Pécora, J.D., Silva-Sousa, Y.T. & de Sousa-Neto, M.D. 2016. Zinc Oxide Nanoparticles Enhance Physicochemical Characteristics of Grossman Sealer. *Journal of Endodontics*. 42(12):1804–1810.
- Vidhya, E., Vijayakumar, S., Prathipkumar, S. & Praseetha, P.K. 2020. Green way biosynthesis: Characterization, antimicrobial and anticancer activity of ZnO nanoparticles. *Gene Reports*. 20:100688.
- Vijayakumar, S., Krishnakumar, C., Arulmozhi, P., Mahadevan, S. & Parameswari, N. 2018a. Biosynthesis, characterization and antimicrobial activities of zinc oxide nanoparticles from leaf extract of *Glycosmis pentaphylla* (Retz.) DC. *Microbial Pathogenesis*. 116:44–48.
- Vijayakumar, S., Mahadevan, S., Arulmozhi, P., Sriram, S. & Praseetha, P.K. 2018b. Green synthesis of zinc oxide nanoparticles using *Atalantia monophylla* leaf extracts: Characterization and antimicrobial analysis. *Materials Science in Semiconductor Processing*. 82:39–45.
- Vijayalakshmi, R. & Kumar, S. 2006. Nanotechnology in dentistry. *Indian Journal of Dental Research*. 17(2):62.
- Vila, T., Sultan, A.S., Montelongo-Jauregui, D. & Jabra-Rizk, M.A. 2020. Oral candidiasis: A disease of opportunity. *Journal of Fungi*. 6(1):15.
- Villanueva-Flores, F., Castro-Lugo, A., Ramírez, O.T. & Palomares, L.A. 2020. Understanding cellular interactions with nanomaterials: towards a rational design of medical nanodevices. *Nanotechnology*. 31(13):132002.
- Vinotha, V., Iswarya, A., Thaya, R., Govindarajan, M., Alharbi, N.S., Kadaikunnan, S., Khaled, J.M., Al-Anbr, M.N., *et al.* 2019. Synthesis of ZnO nanoparticles using insulin-rich leaf extract: Anti-diabetic, antibiofilm and anti-oxidant properties. *Journal of Photochemistry and Photobiology B: Biology*. 197(June):111541.

- Wahab, R., Kim, Y.S., Mishra, A., Yun, S. II & Shin, H.S. 2010. Formation of ZnO Micro-Flowers Prepared via Solution Process and their Antibacterial Activity. *Nanoscale Research Letters*. 5(10):1675–1681.
- Wang, D., Cui, L., Chang, X. & Guan, D. 2020. Biosynthesis and characterization of zinc oxide nanoparticles from *Artemisia annua* and investigate their effect on proliferation, osteogenic differentiation and mineralization in human osteoblast-like MG-63 Cells. *Journal of Photochemistry and Photobiology B: Biology*. 202:111652.
- Wang, J., Du, L., Fu, Y., Jiang, P. & Wang, X. 2019. ZnO nanoparticles inhibit the activity of *Porphyromonas gingivalis* and *Actinomyces naeslundii* and promote the mineralization of the cementum. *BMC Oral Health*. 19(1):1–11.
- Wang, J., Gao, S., Wang, S., Xu, Z. & Wei, L. 2018. Zinc oxide nanoparticles induce toxicity in CAL 27 oral cancer cell lines by activating PINK1/Parkin-mediated mitophagy. *International Journal of Nanomedicine*. 13:3441–3450.
- Wang, L., Hu, C. & Shao, L. 2017. The antimicrobial activity of nanoparticles: Present situation and prospects for the future. *International Journal of Nanomedicine*. 12:1227–1249.
- Wani, A.H. & Shah, M.A. 2012. A unique and profound effect of MgO and ZnO nanoparticles on some plant pathogenic fungi. *Journal of Applied Pharmaceutical Science*. 2012(03):40–44.
- Webb, B.C., Thomas, C.J., Willcox, M.D., Harty, D.W. & Knox, K.W. 1998. Candida-associated denture stomatitis. Aetiology and management: a review. Part 1. Factors influencing distribution of Candida species in the oral cavity. *Australian dental journal*. 43(1):45–50.
- Williams, D.B. & Carter, C.B. 1996. Scattering and Diffraction. in *Transmission Electron Microscopy: A textbook for materials science*. Springer US. 19–33.
- Williams, D.B. & Carter, C.B. 2009. *Transmission electron microscopy: A textbook for materials science*. Second edition. Springer US:1–805
- Witbooi, H., Okem, A., Makunga, N.P. & Kambizi, L. 2017. Micropropagation and secondary metabolites in *Agathosma betulina* (Berg.). *South African Journal of Botany*. 111:283–290.
- Van Wyk, B.E. 2008. A broad review of commercially important southern African medicinal plants. *Journal of Ethnopharmacology*. 119(3):342–355.
- Xie, Y., He, Y., Irwin, P.L., Jin, T. & Shi, X. 2011. Antibacterial activity and mechanism of action of zinc oxide nanoparticles against *Campylobacter jejuni*. *Applied and Environmental Microbiology*. 77(7):2325–2331.
- Yamamoto, O. 2001. Influence of particle size on the antibacterial activity of zinc oxide. *International Journal of Inorganic Materials*. 3(7):643–646.
- Yao, L.H., Jiang, Y.M., Shi, J., Tomás-Barberán, F.A., Datta, N., Singanusong, R. & Chen, S.S. 2004. Flavonoids in food and their health benefits. *Plant Foods for Human Nutrition*. 59(3):113–122.
- Yusof, H.M., Mohamad, R., Zaidan, U.H. & Abdul Rahman, N.A. 2019. Microbial synthesis of zinc oxide nanoparticles and their potential application as an antimicrobial agent and a feed supplement in animal industry: A review. *Journal of Animal Science and Biotechnology*. 10(1):57.

- Yuvakkumar, R., Suresh, J., Nathanael, A.J., Sundrarajan, M. & Hong, S.I. 2014a. Novel green synthetic strategy to prepare ZnO nanocrystals using rambutan (*Nephelium lappaceum L.*) peel extract and its antibacterial applications. *Materials Science and Engineering C*. 41:17–27.
- Yuvakkumar, R., Suresh, J. & Hong, S.I. 2014b. Green Synthesis of Zinc Oxide Nanoparticles. *Advanced Materials Research*. 952(3):137–140.
- Zacchino, S.A., Butassi, E., Liberto, M. Di, Raimondi, M., Postigo, A. & Sortino, M. 2017a. Plant phenolics and terpenoids as adjuvants of antibacterial and antifungal drugs. *Phytomedicine*. 37:27–48.
- Zacchino, S.A., Butassi, E., Cordisco, E. & Svetaz, L.A. 2017b. Hybrid combinations containing natural products and antimicrobial drugs that interfere with bacterial and fungal biofilms. *Phytomedicine*. 37:14–26.
- Zagorac, D., Schön, J., Pentin, V. & Jansen, M. 2011. Structure prediction and energy landscape exploration in the zinc oxide system. *Processing and Application of Ceramics*. 5(2):73–78.
- Zanatta, F.B., Antoniazzi, R.P. & Rösing, C.K. 2010. Staining and calculus formation after 0.12% chlorhexidine rinses in plaque-free and plaque covered surfaces: A randomized trial. *Journal of Applied Oral Science*. 18(5):515–521.
- Zare, E., Pourseyedi, S., Khatami, M. & Darezereshki, E. 2017. Simple biosynthesis of zinc oxide nanoparticles using nature's source, and its *in vitro* bio-activity. *Journal of Molecular Structure*. 1146:96–103.
- Zhang, H. & Chen, G. 2009. potent antibacterial activities of Ag/TiO₂ nanocomposite powders synthesized by a one-pot sol-gel method. *Environmental Science and Technology*. 43(8):2905–2910.
- Zhou, W. & Wang, Z.L. 2007. Scanning microscopy for nanotechnology: Techniques and applications. Springer New York:1–513.

APPENDIX

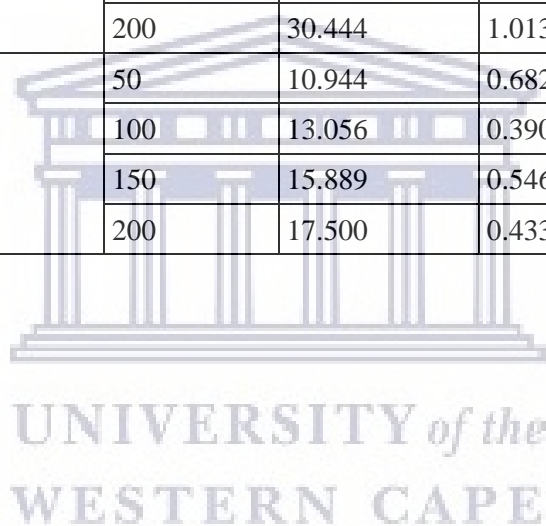
Supplemental tables

Supplemental Table 1: Actual measurements of inhibition zones diameters from the Kirby Bauer assay for the three intervention groups [CHX, GZnO NPs, and Zn (NO₃)₂·6H₂O solution]

Intervention	Volume (µL)	Zones of inhibition (diameter in mm)									Average (mm)
0.2% Chlorhexidine (CHX) gluconate	50	10	11	11	12	11	11	11	11.5	11.5	11.11
	100	22	22	22	22	22	22	22	22	21.5	21.94
	150	23.5	24	23.5	23	23.5	23	24	23.5	23.5	23.5
	200	28	27	28	26.5	27	26.5	28	26.5	26.5	27.11
Zinc oxide nanoparticles (GZnO NPs)	50	12	13	14	13	12.5	13	13	13	13	12.94
	100	21	21	22	20	20	19	21	20	21	20.55
	150	22	23	24	22.5	23	23	23	23	23.5	23
	200	30	30	32	31	32	30	29	30	30	30.44
Zinc nitrate hexahydrate [Zn (NO₃)₂·6H₂O] solution	50	10.5	11	10	12	11	12	11	10.5	10.5	10.94
	100	13	13	12.5	13	13	13	14	13	13	13.05
	150	16	17	16	15.5	16	15	16	16	15.5	15.88
	200	18	17.5	17	17.5	18	17	18	17	17.5	17.5

Supplemental Table 2: The mean diameters of inhibition zone and their respective standard deviations using the Kirby Bauer assay for the three intervention groups

Intervention	Volume (μL)	Zones of inhibition (mean) in mm	Standard deviation	n
0.2% Chlorhexidine (CHX) gluconate	50	11.111	0.5465	9
	100	21.944	0.1667	9
	150	23.500	0.3536	9
	200	27.111	0.6972	9
Zinc oxide nanoparticles (GZnO NPs)	50	12.944	0.5270	9
	100	20.556	0.8819	9
	150	23.000	0.5590	9
	200	30.444	1.0138	9
Zinc nitrate hexahydrate [Zn (NO₃)₂·6H₂O] solution	50	10.944	0.6821	9
	100	13.056	0.3909	9
	150	15.889	0.5465	9
	200	17.500	0.4330	9



Supplemental Table 3: The mean optical densities with their respective standard deviations using the CV staining assay for the three intervention groups

Intervention (100 µL)	Time (hours)	Mean for optical densities (560 nm)	Standard deviation	n
0.2% Chlorhexidine (CHX) gluconate	0	2.3442	0.143	28
	2	2.1187	0.256	28
	4	2.2440	0.017	28
	6	2.2677	0.041	28
	8	2.4592	0.058	28
	12	2.3781	0.168	28
	24	2.2864	0.101	28
	48	2.1821	0.068	28
	72	1.7611	0.188	28
Zinc Oxide Nanoparticles (GZnO NPs)	0	1.5578	0.259	28
	2	1.7072	0.467	28
	4	1.7431	0.043	28
	6	1.6815	0.019	28
	8	1.7851	0.201	28
	12	1.7638	0.100	28
	24	1.5206	0.437	28
	48	1.3513	0.313	28
	72	0.7912	0.107	28
Nystatin	0	2.1380	0.280	28
	2	1.8502	0.128	28
	4	1.8169	0.108	28
	6	1.5127	0.031	28
	8	1.4785	0.380	28
	12	2.1979	0.220	28
	24	1.8236	0.101	28
	48	1.5952	0.113	28
	72	1.3211	0.222	28

Supplemental Table 4: The confidence levels of the multiple comparisons (Tukey's HSD tests) using CV staining assay for the three intervention groups at different time points

Time (hours)	Intervention (I)	Intervention(J)	Mean difference (I-J) of optical density	Significance (P-value)	95% Confidence Interval	
					Lower Bound	Upper Bound
0	GZnO NPs	Chlorhexidine	0.08252	0.7300	-0.1291	0.2942
	GZnO NPs	Nystatin	-0.02783	0.9852	-0.2395	0.1838
	GZnO NPs	Negative control	0.2390	0.0211*	0.02730	0.4506
	CHX	Nystatin	-0.1104	0.5151	0.3220	0.1013
	CHX	Negative control	0.1564	0.2157	-0.05522	0.3681
	Nystatin	Negative control	0.2668	0.0081**	0.05513	0.4785
2	GZnO NPs	Chlorhexidine	0.3539	<.0001****	0.2225	0.4853
	GZnO NPs	Nystatin	0.3111	<.0100****	0.1797	0.4425
	ZnO NPs	Negative control	0.2070	0.0006****	0.07563	0.3384
	CHX	Nystatin	-0.04279	0.8230	-0.1742	0.08858
	CHX	Negative control	-0.1469	0.0228*	-0.2783	-0.01552
	Nystatin	Negative control	-0.1041	0.1656	-0.2355	0.02727
4	GZnO NPs	Chlorhexidine	0.3141	0.0076**	0.06656	0.5616
	GZnO NPs	Nystatin	0.3677	0.0013**	0.1202	0.6152
	GZnO NPs	Negative control	0.6684	<.0100****	0.4208	0.9159
	CHX	Nystatin	0.05361	0.9392	-0.1939	0.3011
	CHX	Negative control	0.3543	0.0021**	0.1067	0.6018
	Nystatin	Negative control	0.3007	0.0114*	0.05311	0.5482
6	GZnO NPs	Chlorhexidine	0.2427	0.0032**	0.06659	0.4188
	GZnO NPs	Nystatin	0.5210	<.0001****	0.3449	0.6971
	GZnO NPs	Negative control	0.5752	<.0001****	0.3991	0.7513
	CHX	Nystatin	0.2783	0.0006****	0.1022	0.4544
	CHX	Negative control	0.3325	<.0001****	0.1564	0.5086
	Nystatin	Negative control	0.05426	0.8458	-0.1218	0.2304
8	GZnO NPs	Chlorhexidine	0.2274	<.0001****	0.1201	0.3347
	GZnO NPs	Nystatin	0.64143066	<.0001****	0.5341	0.7488
	GZnO NPs	Negative control	0.6494	<.0001****	0.5421	0.7567

Time (hours)	Intervention (I)	Intervention(J)	Mean difference (I-J) of optical density	Significance (P-value)	95% Confidence Interval	
					Lower Bound	Upper Bound
	CHX	Nystatin	0.4140	<.0001****	0.3067	0.5214
	CHX	Negative control	0.4220	<.0001****	0.3147	0.5294
	Nystatin	Negative control	0.007989	0.9972	-0.09934	0.1153
12	GZnO NPs	Chlorhexidine	0.2549	<.0001****	0.1515	0.3584
	GZnO NPs	Nystatin	0.1229	<.0138*	0.01946	0.2264
	GZnO NPs	Negative control	0.1172	0.0205*	0.01374	0.2207
	CHX	Nystatin	-0.1320	0.0072**	-0.2355	-0.02857
	CHX	Negative control	-0.1377	0.0047**	-0.2412	-0.03429
	Nystatin	Negative control	-0.005719	0.9989	-0.1092	0.09774
24	GZnO NPs	Chlorhexidine	0.08156	0.0919	-0.009060	0.1722
	GZnO NPs	Nystatin	0.1553	0.0002****	0.06470	0.2459
	GZnO NPs	Negative control	0.1587	0.0001***	0.06808	0.2493
	CHX	Nystatin	0.07376	0.1481	-0.01686	0.1644
	CHX	Negative control	0.07714	0.1211	-0.01348	0.1678
	Nystatin	Negative control	0.003375	0.9996	-0.08725	0.09400
48	GZnO NPs	Chlorhexidine	-0.01543	0.9906	-0.1524	0.1216
	GZnO NPs	Nystatin	0.1556	0.0202*	0.01857	0.2926
	GZnO NPs	Negative control	0.1212	0.1003	-0.01579	0.2583
	CHX	Nystatin	0.1710	0.0089**	0.03400	0.3080
	CHX	Negative control	0.1367	0.0508	-0.0003604	0.2737
	Nystatin	Negative control	-0.03436	0.9096	-0.1714	0.1027
72	GZnO NPs	Chlorhexidine	-0.2918	<.0001****	-1.4367	-0.1470
	GZnO NPs	Nystatin	-0.1772	0.0107*	-0.3220	-0.03231
	GZnO NPs	Negative control	-0.08054	0.4593	-0.2254	0.06433
	CHX	Nystatin	0.1146	0.1665	-0.03021	0.2595
	CHX	Negative control	0.2113	0.0017**	-0.06642	0.3561
	Nystatin	Negative control	0.09663	0.2990	-0.04823	0.2415

Supplemental Table 5: Means optical density for biofilm reduction of *C. albicans* for the three interventions at consecutive time point across the 72 Hours using CV staining assay

Average gradients for each intervention	2hr-0hr	12hr-2hr	24hr-12hr	48hr-24hr	72hr-48hr	MEANS	STDEV
CHX	-0.113	0.026	-0.008	-0.004	-0.018	-0.023	0.047
Nystatin	-0.144	0.035	-0.031	-0.010	-0.011	-0.032	0.060
GZnO NPs	0.075	0.006	-0.020	-0.007	-0.023	0.006	0.036
Neg. Control	0.086	0.014	-0.022	-0.005	-0.013	0.012	0.039

